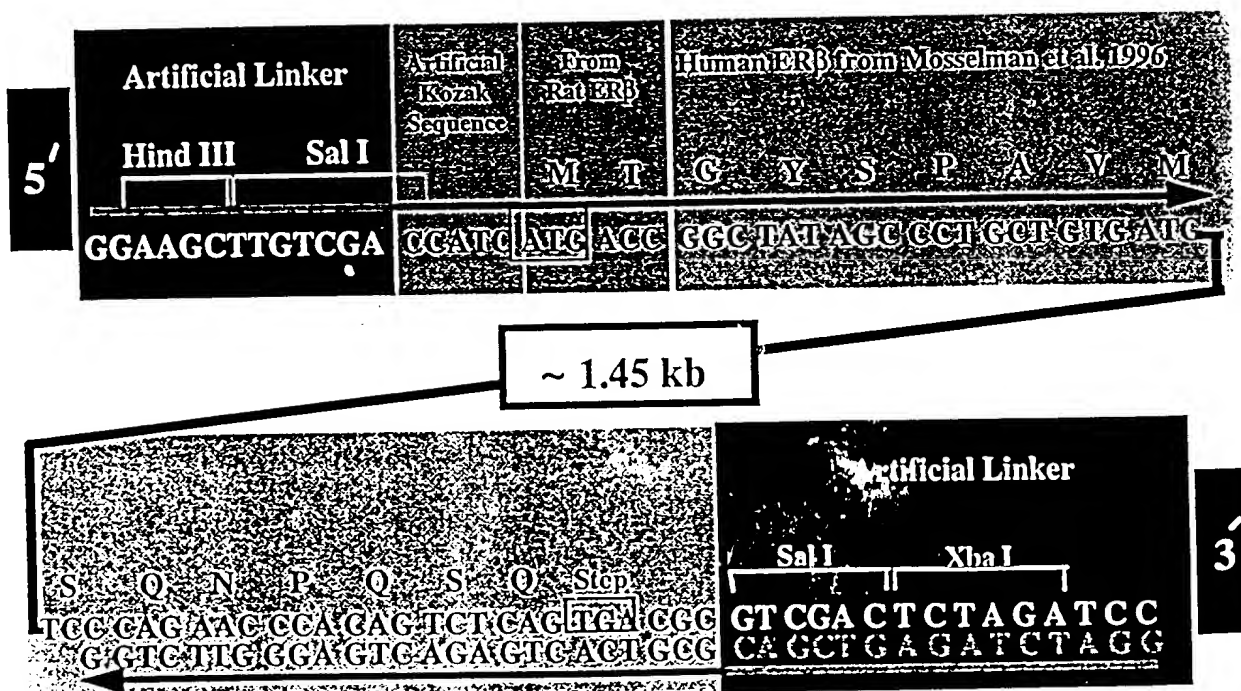


FIGURE 1

## PCR Cloning of the Human ER $\beta$

Human testis RNA was reverse transcribed using Oligo dT. The resulted transcript was used for PCR with Oligonucleotides (red arrows) designed as follows:



The PCR product was cloned in the Hind III and XbaI sites of the Eukaryotic expression vector pcDNA III.

FIGURE 2

# Human ER $\beta$ cDNA

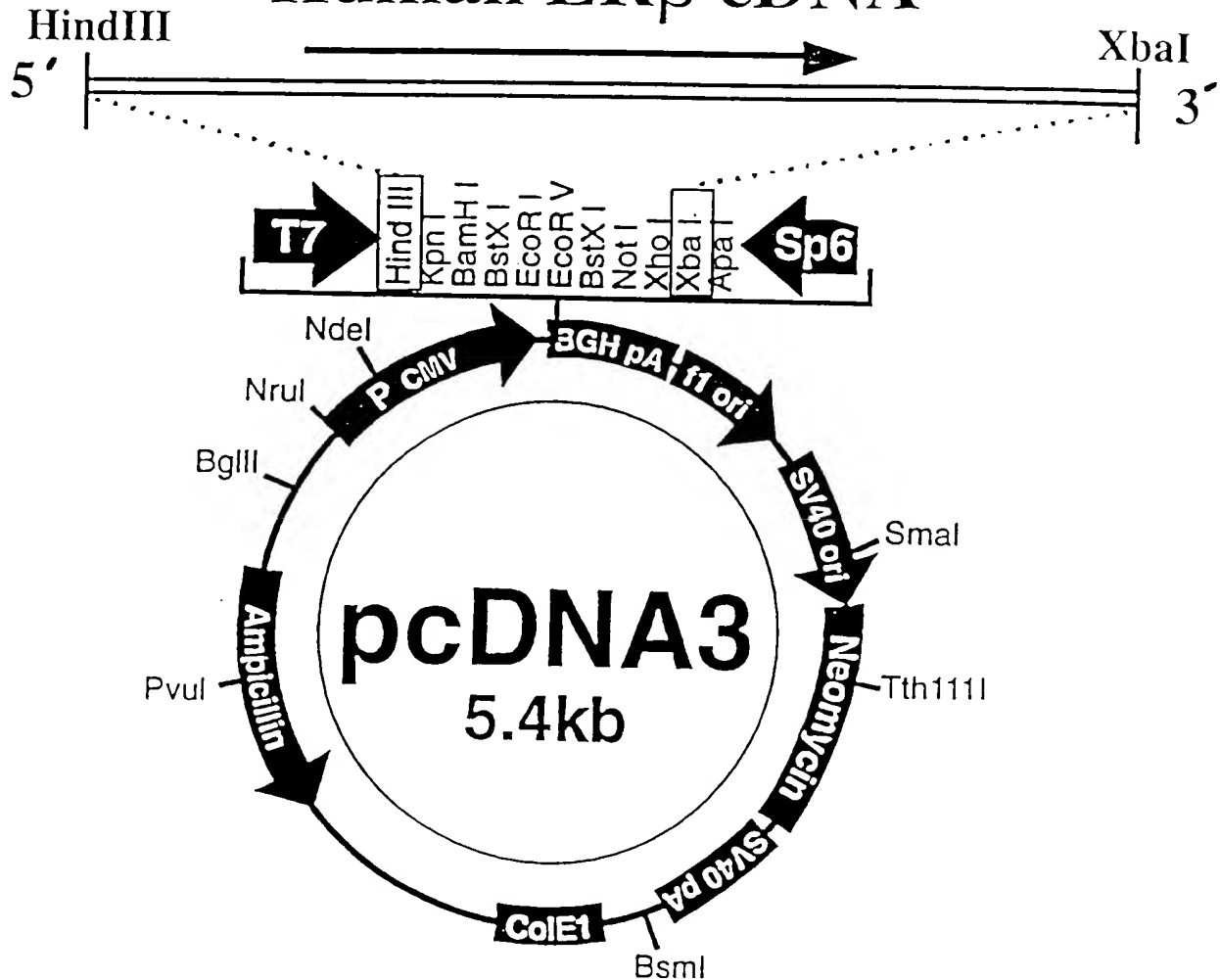


Fig 2

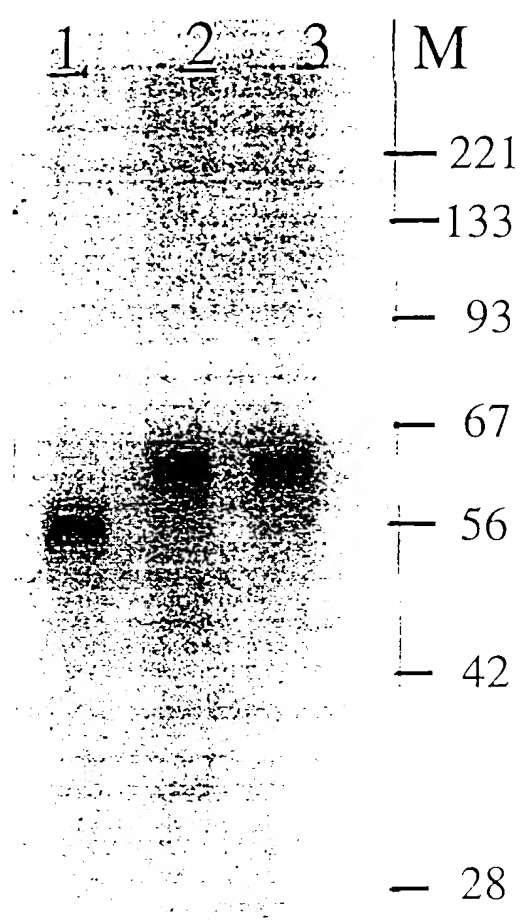
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 TGCAAGGTGTTTTCTCAGCTGCTATCTCAAGACATGGATATAAAAACTCACCATCTAGC 120  
 CTTAATTCTOCTTCTOCTCTACAACTGCAGTCAATCCATCTTACCCCTGGAGCAOCCGCTOC 180  
 ATATACATAOCTTCTOCTCTATG TAGACAGOCACCATGAATATCCAGCCATGACATTCTAT 240  
 AGOCCCTGCTGTGATGAATTACAGCATTOCCAGCAATGTCACTAACTTGGAAGGTGGGOCT 300  
 GGTGCGGCAGAACACAAGOOCAAATGTGTTGTGGOCAACAOCCTGGGCAOCTTTCTOCTTTA 360  
 GTGGTCCATOGOCAGTTATCACATCTGTATGOGGAOCTCAAAAGAGTOOCTGGTGTGAA 420  
 GCAAGATOGCTAGAACACAOCCTTAOCTGTAAACAGAGAGACACTGAAAAGGAAGGTTAGT 480  
 GGGAAOOGTTGOGOCAGOOCTGTTACTGGTOCAGGTTCAAAGAGGGATGCTCACTTCTGC 540  
 GCTGTCTGCAGOGATTACGCATOGGGATATCACTATGGAGTCTGGTCTGTGAAGGATGT 600  
 AAGGCCTTTTTTAAAAAGAAGCATTCAAGGACATAATGATTATATTTGTCCAGCTACAAAT 660  
 CAGTGTACAATOGATAAAAAOOGGOGCAAGAGCTGOCAGGOCTGOOGACTTOGGAAGTGT 720  
 TAOGAAGTGGGAATGGTGAAGTGTGGCTOOOGGAGAGAGAGATGTGGGTACOCGCTTGTG 780  
 OGGAGACAGAGAAGTGOOGAOCAGCAGCTGCACTGTGOOGGCAAGGCCAAGAGAAGTGGC 840  
 GGOCACOGOGOOOGAGTGOGGGAGCTGCTGCTGGAOGOOCTGAGOOOOGAGCAGCTAGTG 900  
 CTCACCCCTOCTGGAGGCTGAGOOOGOOOCATGTGCTGATCAGOOOGOOOCAGTGOGOOCTTC 960  
 ACOGAGGOCTCCATGATGATGTTOCTGAOCCAAGTTGGOOGACAAGGAGTTGGTACACATG 1020  
 ATCAGCTGGGGOCAAGAAGATTCCOOGGCTTTGTGGAGCTCAGOCTGTTGACCAAGTGOGG 1080  
 CTCTTGGAGAGCTGTTGGATGGAGGTGTTAATGATGGGGCTGATGTGGOGCTCAATTGAC 1140  
 CACCCOOGGCAAGCTCATCTTTGCTCCAGATCTTGTTCTGGACAGGGATGAGGGGAAATGC 1200  
 GTAGAAGGAATTCTGGAAATCTTTGACATGCTCCTGGCAACTACTTCAAGGTTTCCAGAG 1260  
 TTAAAACTCCAACACAAAGAATATCTCTGTGTCAAGGOCATGATCCTGCTCAATTCCAGT 1320  
 ATGTACCCCTCTGGTCACAGOGAOCAGGATGCTGACAGCAGOOGGAAGCTGGCTCACTTG 1380  
 CTGAAOOGOGTGACOGATGCTTTGGTTTGGGTGATTGCCAAGAGOGGCATCTOCTCCOCAG 1440  
 CAGCAATOCATGOGCTGGCTAAOCTOCTGATGCTOCTGTCCACGTCAGGCATGOGAGT 1500  
 AACAAGGGCATGGAACATCTGCTCAACATGAAGTGCAAAAATGTGGTCCAGTGTATGAC 1560  
 CTGCTGCTGGAGATGCTGAATGOCCAOGTGCTTOGOGGGTGCAAGTOCTOCATCAOGGGG 1620  
 TCOGAGTGACGOOOGGCAGAGGACAGTAAAAGCAAAGAGGGCTOCCAGAAOCCACAGTCT 1680  
 CAGTGA 1686

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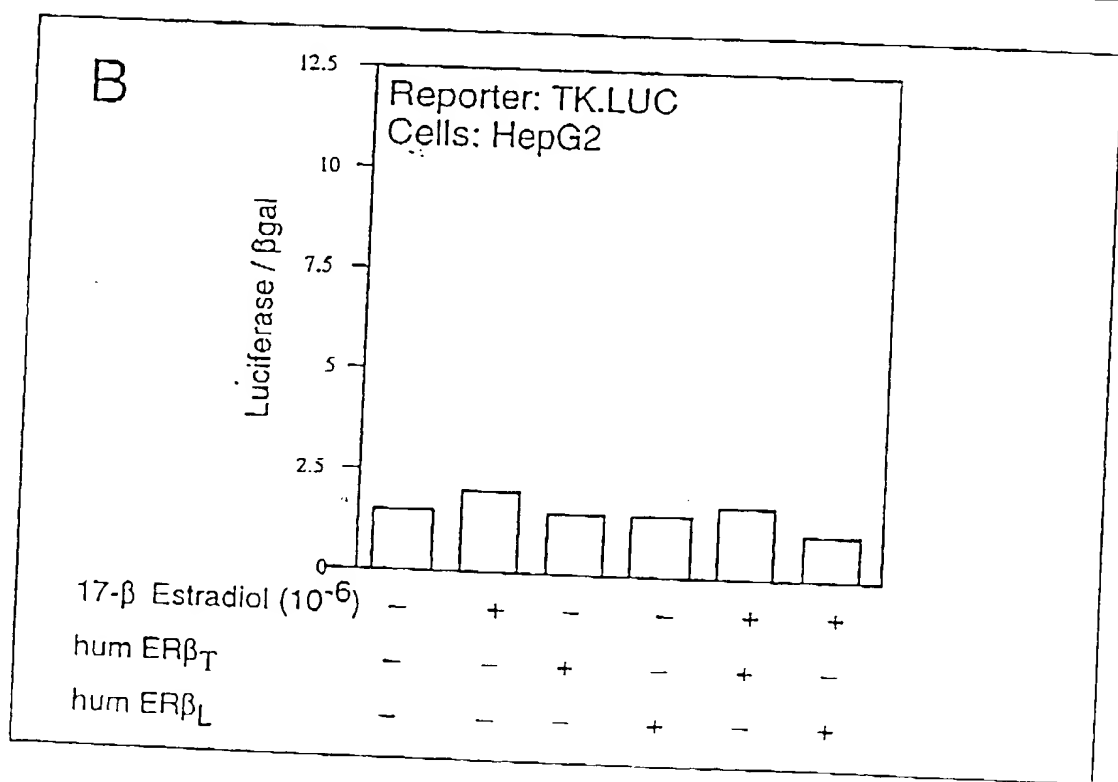
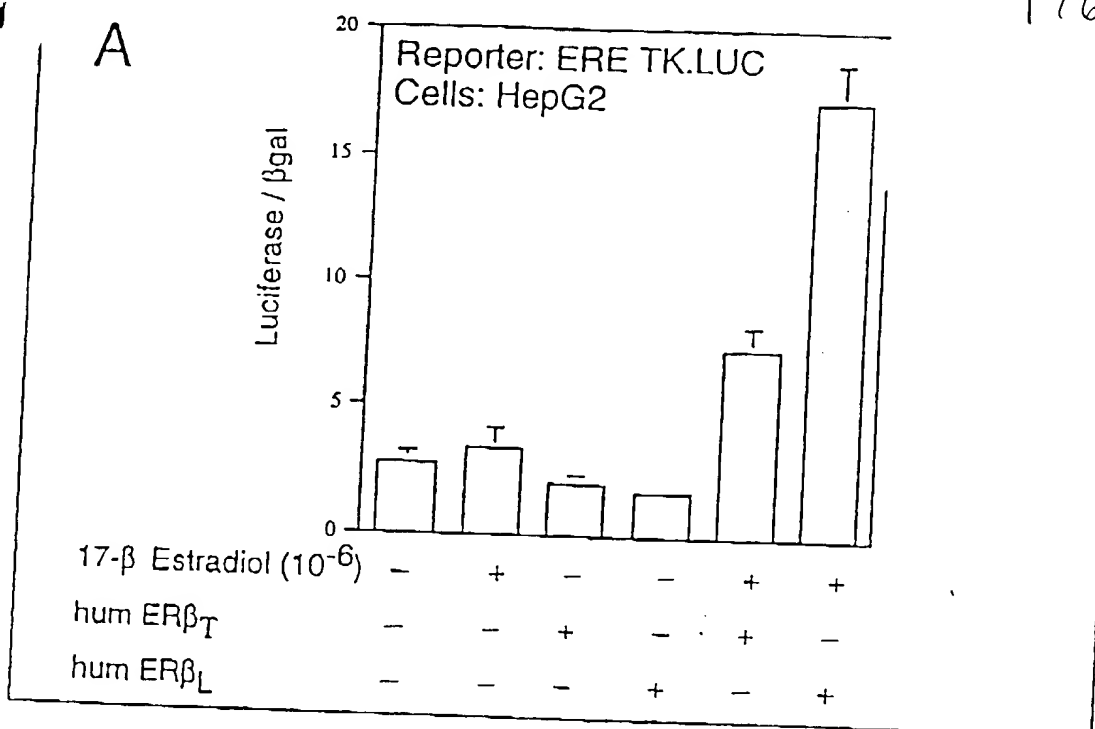
<u>MDIKNSPSSL</u>	<u>NSPSSYNCSO</u>	<u>SILPLEHCSI</u>	<u>YIPSSYVDSH</u>	<u>HEYPAMTFYS</u>	50
PAVMNYSIPS	NVINLEGGPG	RQTTSPNVLW	PTPGHLSPLV	VHRQLSHLYA	100
EPQKSPWCEA	RSLEHTLPVN	RETLKRKVS	NRCA SPVTGP	GSKRDAHFCA	150
VCSDYASGYH	YGVWSCEGCK	AFFKRSIQGH	NDYICPATNQ	CTIDKNRRKS	200
CQACRLRKCY	EVGMVKCGSR	REROGYRLVR	RQSADEQLH	CAGKAKRSGG	250
HAPRVRELLL	DALSPEQLVL	TLLEAEPPHV	LISRPSAPFT	EASMMMSLTK	300
LADKELVHMI	SWAKKIPGFV	ELSLFDQVRL	LESCWMEVLM	MGLMWRSIDH	350
PGKLIFAPDL	VLDRDEGKCV	EGILEIFDML	LATTSRFREL	KLQHKEYLCV	400
KAMILLNSSM	YPLVTATQDA	DSSRKLALL	NAVTDALVW	IAKSGISSQQ	450
QSMRLANLLM	LLSHVRHASN	KGMEHLLNMK	CKNVVPVYDL	LLEMLNAHVL	500
RGCKSSITGS	ECSPAEDSKS	KEGSONPQSQ	.	531	

1764

Fig 5



F766



Transactivation of ERE reporter By ER $\beta_L$  and ER $\beta_T$  In HepG2 cells. Luciferase reporter constructs (0.5 ug) containing either [A] the estrogen receptor DNA response element upstream of the TK basal promoter (ERE TK.LUC) or [B] the TK basal promoter alone (TK.LUC) were transiently transfected into HepG2 cells by the calcium phosphate coprecipitation method. Each construct was cotransfected with the ER expression vector (0.25 ug) indicated and the RSV- $\beta$ -galactosidase plasmid (0.5 ug) to correct for variation in DNA uptake. Luciferase activity was normalized to  $\beta$ -galactosidase enzymatic activity.

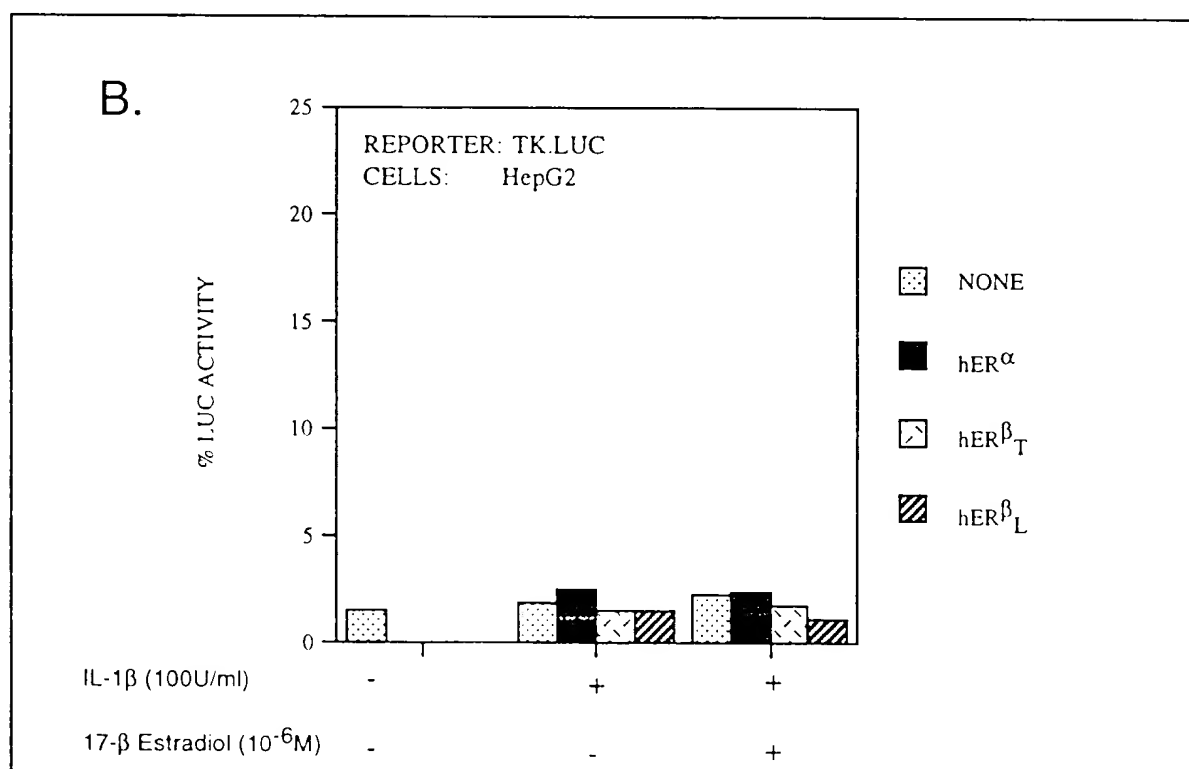
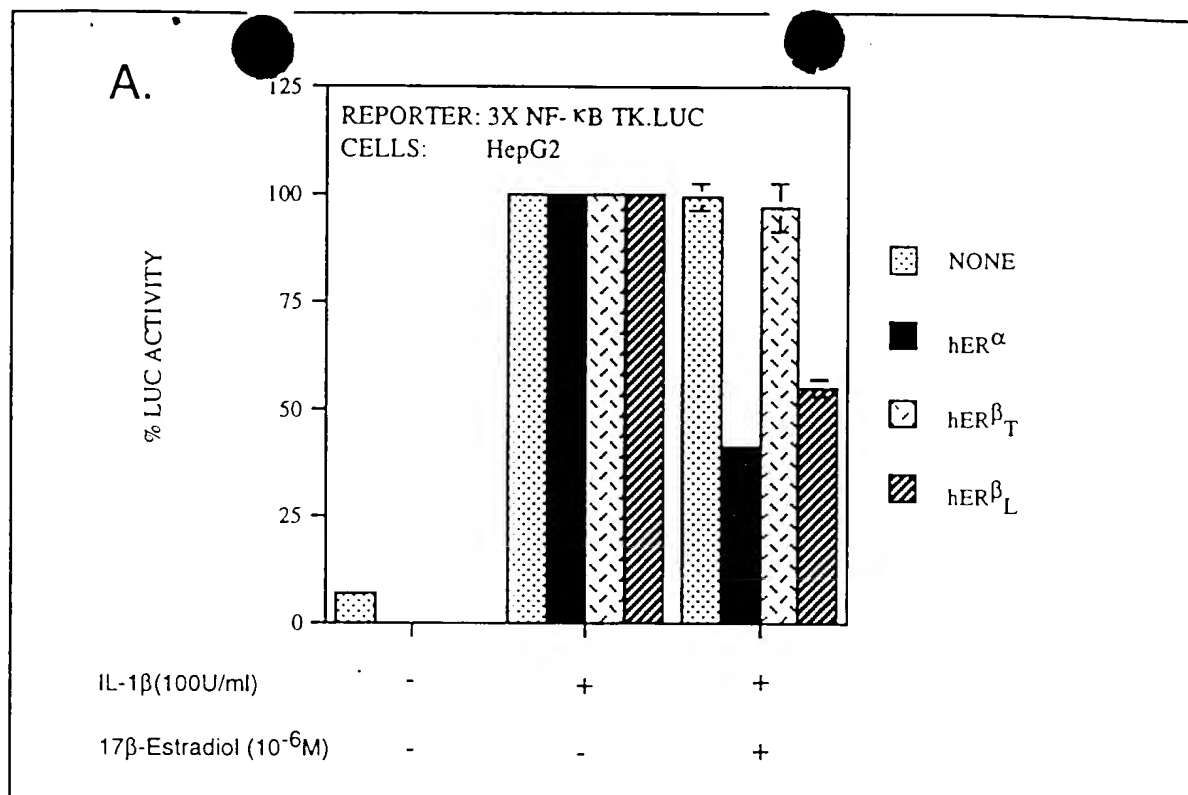
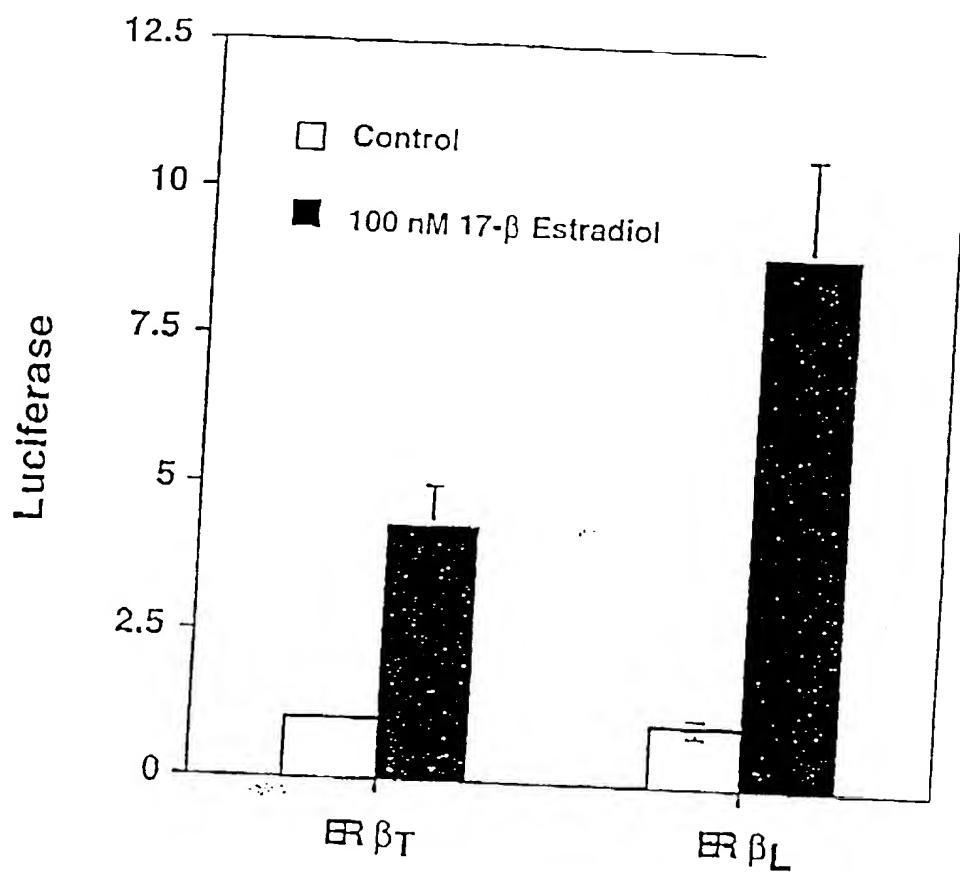


Fig 7

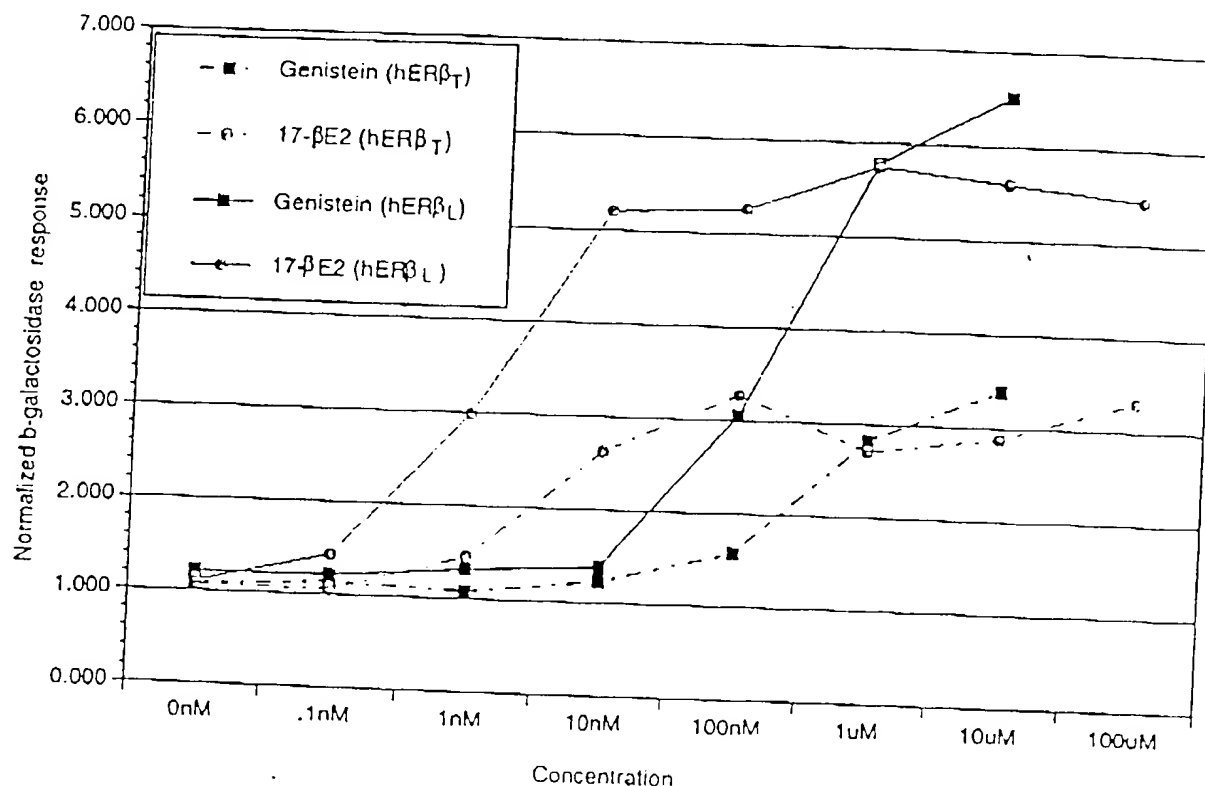
Luciferase reporter constructs (0.5  $\mu$ g) containing either [A] 3 copies of an NF $\kappa$ B binding site upstream of the TK basal promoter (3X-NF $\kappa$ B TK.LUC) or [B] the TK basal promoter construct alone (TK.LUC) were transiently cotransfected into HepG2 cells by the calcium phosphate coprecipitation method. Each construct was cotransfected with the ER expression vector indicated and the plasmid RSV- $\beta$ -galactosidase (0.5  $\mu$ g) to correct for variation in DNA uptake. Percent luciferase activity values represent Luc: $\beta$ -galactosidase enzymatic activity ratios relative to a value of 100% designated for the IL-1 $\beta$  treated samples and are presented as mean  $\pm$  S.E..



Transactivation of ERE reporter by ERβ<sub>T</sub> and ERβ<sub>L</sub> in HAECT-1 cells. Luciferase reporter constructs (20 μg) containing either the estrogen receptor DNA response element upstream of the TK basal promoter (ERE TK.LUC) or the TK basal promoter (TK.Luc) were transiently transfected into HAECT-1 cells (4x10<sup>6</sup>) with 5 μg of ER expression vector by electroporation. Cells were plated into 48 wells of a 96-well plate, rested for 4h, and treated overnight as indicated prior to luciferase determination. ERE TK.LUC values were normalized to TK.LUC values and are presented as mean ± S.E. (n=4).



K69



Transcriptional activity of hERβ<sub>T</sub> and hERβ<sub>L</sub> in yeast. Yeast cells (BJ2168) were cotransformed with an ERE-LacZ reporter (YRpE2) and either a yeast vector (pYX242) expressing hERβ<sub>T</sub> or hERβ<sub>L</sub>. Transformed cells were grown in selective medium for 24 h at 30°C. Cells were treated with 17-β estradiol or genistein, at the indicated concentrations, for 3 h and then assayed for β-galactosidase activity.